



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁵ : C12Q 1/00, 1/68	A1	(11) International Publication Number: WO 90/12111 (43) International Publication Date: 18 October 1990 (18.10.90)
(21) International Application Number: PCT/US90/01938 (22) International Filing Date: 10 April 1990 (10.04.90) (30) Priority data: 336,751 12 April 1989 (12.04.89) US (71) Applicant: PRESIDENT AND FELLOWS OF HARVARD COLLEGE [US/US]; 17 Quincy Street, Cambridge, MA 02138 (US). (72) Inventors: TABOR, Stanley ; 37 Fayerweather Street, Cambridge, MA 02138 (US). RICHARDSON, Charles, C. ; 78 Chestnut Hill Road, Chestnut Hill, MA 02167 (US). (74) Agent: FRENCH, Timothy, A.; Fish & Richardson, One Financial Center, Suite 2500, Boston, MA 02111-2658 (US).		(81) Designated States: AT (European patent), AU, BB, BE + (European patent), BR, CA, CH (European patent), DE (European patent), DK (European patent), ES (European patent), FI, FR (European patent), GB (European patent), HU, IT (European patent), JP, KP, KR, LK, LU (European patent), MC, MG, MW, NL (European patent), NO, RO, SD, SE (European patent), SU. Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
(54) Title: IMPROVED PRIMER EXTENSION REACTIONS (57) Abstract A kit or solution for use in extension of an oligonucleotide primer having a first single-stranded region on a template molecule having a second single-stranded region homologous to the first single-stranded region, comprising a first agent able to cause extension of the first single-stranded region of the primer on the second single-stranded region of the template in a reaction mixture, and a second agent able to reduce the amount of pyrophosphate in the reaction mixture below the amount produced during the extension in the absence of the second agent.		

DESIGNATIONS OF "DE"

Until further notice, any designation of "DE" in any international application whose international filing date is prior to October 3, 1990, shall have effect in the territory of the Federal Republic of Germany with the exception of the territory of the former German Democratic Republic.

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	ES	Spain	MG	Madagascar
AU	Australia	FI	Finland	ML	Mali
BB	Barbados	FR	France	MR	Mauritania
BE	Belgium	GA	Gabon	MW	Malawi
BF	Burkina Faso	GB	United Kingdom	NL	Netherlands
BG	Bulgaria	HU	Hungary	NO	Norway
BJ	Benin	IT	Italy	RO	Romania
BR	Brazil	JP	Japan	SD	Sudan
CA	Canada	KP	Democratic People's Republic of Korea	SE	Sweden
CF	Central African Republic	KR	Republic of Korea	SN	Senegal
CG	Congo	LI	Liechtenstein	SU	Soviet Union
CH	Switzerland	LK	Sri Lanka	TD	Chad
CM	Cameroon	LJ	Luxembourg	TG	Togo
DE	Germany, Federal Republic of	MC	Monaco	US	United States of America
DK	Denmark				

IMPROVED PRIMER EXTENSION REACTIONS

Background of the Invention

This invention was made with government support including a grant from Department of Energy Grant No. DE-SG02-88ER60688 and U.S. Public Health Service Grant No. Al-06045. The U.S. government has certain rights to the invention.

This application is a continuation-in-part of Tabor et al., entitled DNA SEQUENCING, U.S. Serial No. 218,103, filed July 12, 1988, which is hereby incorporated by reference herein.

This invention relates to methods for performing a primer extension reaction, such as a DNA sequencing reaction, or a polymerase chain reaction.

In a primer extension reaction an oligonucleotide primer having homology to a single-stranded template DNA, e.g., genomic DNA, is caused to anneal to the template DNA. The annealed mixture is then provided with a DNA polymerase in the presence of nucleoside triphosphates under conditions in which the DNA polymerase extends the primer to form a complementary DNA strand to the template DNA. In a DNA sequencing reaction, the primer is extended in the presence of a chain-terminating agent, e.g., a dideoxynucleoside triphosphate, to cause base-specific termination of the primer extension. Sanger et al., 74 Proc. Nat'l. Acad. Sci. 5463, 1977. In a polymerase chain reaction two primers are provided, each having homology to opposite strands of a double-stranded DNA molecule. After the primers are extended, they are separated from their templates, and additional primers caused to anneal to the templates and the extended primers. The additional primers are then extended. The steps of separating, annealing, and extending are

- 2 -

repeated in order to amplify the number of copies of template DNA. Saiki et al., 239 Science 487, 1988.

Summary of the Invention

5 In a first aspect, the invention features a solution or kit for use in extension of an oligonucleotide primer having a first single-stranded region on a template molecule having a second single-stranded region, the first and second regions being homologous. The solution or kit includes a first agent able to cause extension of the first single
10 stranded region of the primer on the second single-stranded region of the template in a reaction mixture, and a second agent able to reduce the level of pyrophosphate in the reaction mixture below the level
15 produced during extension in the absence of the second agent.

By solution is meant any aqueous and/or buffered liquid containing the components described above. These components are present in the solution at
20 concentrations sufficient to perform their desired function. For example, the first agent is present at a concentration sufficient to reduce the level of pyrophosphate in the solution. By kit is meant a container which holds one or more of the components of
25 the solution separately. For example, the first and second agents are held in separate containers in solutions adapted to be mixed together.

By causing extension of the oligonucleotide primer is meant performing a reaction in which an
30 oligonucleotide primer having a single-stranded region is annealed, or naturally occurs in the annealed state, with another nucleic acid molecule which acts as a template upon which the oligonucleotide primer can be

- 3 -

extended by addition of nucleoside triphosphates to form nucleic acid homologous to the template nucleic acid. Generally, extension entails providing a DNA polymerase or RNA polymerase to covalently add nucleotides to the primer.

A reaction mixture is any solution or solid phase suitable for performing an extension reaction. Generally, it is a liquid buffer containing nucleoside or deoxynucleoside triphosphates and metal ions required for an extension reaction. The mixture may also contain any standard buffering agents and, for a DNA sequencing reaction, one or more dideoxynucleoside triphosphates, or an equivalent chain-terminating agent.

By reducing the level of pyrophosphate is meant that the amount of pyrophosphate in the reaction mixture is reduced to an amount which has little or no significant effect on the extension of the primer on the template. That is, the level of pyrophosphate is low enough to reduce pyrophosphorolysis to an insignificant level (less than 10% the level of pyrophosphorolysis in the presence of 300 μ M pyrophosphate). Preferably, the level of pyrophosphate is reduced to below 25 μ M, even more preferably to below 5 μ M. This phase is meant to include use of an agent, such as a pyrophosphatase, which acts to prevent the build-up of pyrophosphate, as well as remove it from a solution.

By homologous is meant that the two single-stranded regions are able to form sufficient non-covalent bonds between their respective nucleotides to form a stable double-stranded structure under conditions normally used for annealing nucleic acids, and for performing a primer extension reaction.

- 4 -

In preferred embodiments, the first agent is a DNA polymerase, most preferably chosen from Klenow, Taq polymerase, a T7-type DNA polymerase (i.e., a polymerase similar to that in a phage in which the DNA polymerase requires host thioredoxin as a subunit, e.g., T7 DNA polymerase or the DNA polymerase of T3, Φ I, Φ II, H, W31, gh-1, Y, AA1122, or Sp6), T4 DNA polymerase, T5 DNA polymerase, Φ 29 DNA polymerase and reverse transcriptase; the second agent is an enzyme, most preferably a pyrophosphatase, for example, a pyrophosphatase resistant to heating at between 60°C and 95°C.

In a second aspect, the invention features an improved method for extending an oligonucleotide primer having a first single-stranded region on a template molecule having a second single-stranded region, including providing a first agent able to cause extension of the primer on the template. The improvement is provision of a second agent able to reduce the amount of pyrophosphate below the amount produced during extension in the absence of the second agent.

In preferred embodiments, the method includes the steps of providing at least one or two oligonucleotide primers having single-stranded regions and at least one or two template molecules having single-stranded regions, and annealing the single-stranded regions of the primers and the templates to form an annealed mixture. The resulting annealed mixture is provided with the first and second agents to cause extension of the primers. The annealed mixture may also be provided with a dideoxynucleoside triphosphate. The method may further include the step

- 5 -

of separating the primers from the templates after their extension, and repeating the steps of providing primers, extending the primers, and separating the primers.

In a related aspect, the invention features a method for amplifying DNA, including performing a polymerase chain reaction in the presence of an agent able to reduce the amount of pyrophosphate in the reaction below the amount produced during a polymerase chain reaction in the absence of the agent. Preferably, the agent is a pyrophosphatase.

In another related aspect, the invention features a method for amplifying DNA including providing a solution of $\Phi 29$ DNA polymerase, a DNA to be amplified, and an agent able to reduce the amount of pyrophosphate in the solution below that amount produced in the absence of the agent.

Applicants have determined that pyrophosphorolysis, where an oligonucleotide chain is reduced in length, is detrimental to a primer extension reaction. The pyrophosphorolysis is caused by the availability of pyrophosphate. For example, a polymerase chain reaction, as described by Cetus (European Patent Application 0,258,017) and by Saiki et al., 239 Science 487, 1988, is inhibited by addition of pyrophosphate even at very low concentrations. This pyrophosphorolysis can be prevented by providing an agent, for example, a pyrophosphatase, capable of removing pyrophosphate. Addition of pyrophosphatase to a polymerase chain reaction greatly enhances the progress of that reaction, and provides superior results compared to use of the method without a pyrophosphatase. Similarly addition of a pyrophosphatase to a DNA sequencing reaction provides more uniformity in

- 6 -

intensities of bands formed in a polyacrylamide gel used to identify products of the sequencing reaction. This uniformity is due to prevention of degradation of specific DNA products by pyrophosphorolysis.

5 Other features and advantages of the invention will be apparent from the following description of the preferred embodiment thereof, and from the claims.

Description of the Preferred Embodiments

10 Any agent which is capable of inhibiting a pyrophosphorolysis reaction is useful in this invention. One way to inhibit pyrophosphorolysis is to break down any pyrophosphate that is generated during a polymerase reaction, by adding the enzyme
15 pyrophosphatase. Even trace addition of a pyrophosphatase (one thousandth the molar ratio of DNA polymerase molecules in a solution) to a primer extension reaction completely stabilizes oligonucleotide fragments produced in a polymerase reaction, by
20 preventing pyrophosphorolysis. The agent should be added at a concentration sufficient to either catalyze the hydrolysis of pyrophosphate in the reaction mixture at a rate that will prevent accumulation of
25 pyrophosphate to a level that will lead to pyrophosphorolysis, or prevent accumulation of pyrophosphate in any other manner. The amount of agent needed is readily determined by standard techniques.

30 There follows an example of the use of pyrophosphatase in a polymerase chain reaction. This example is not limiting to this invention; those skilled in the art will recognize that any primer extension reaction will be benefited by the addition of an agent as described above. Similarly, the use of pyrophosphatase in the examples below is not limiting to

- 7 -

this invention, other agents suitable for reducing the effect of excess pyrophosphate in a primer extension reaction are readily identified by those skilled in the art. The relative concentrations of primer, DNA polymerase, and pyrophosphatase suitable in the invention are readily determined by routine experimentation, and are well known to those in the art.

It is preferable that a pyrophosphatase used in this invention be resistant to heating at high temperatures, since high temperatures are used in a polymerase chain reaction, for example, temperatures between 95°C to 100°C, although temperatures between 65°C and 95°C are also commonly used. Thus, it is advantageous to provide a pyrophosphate resistant to heating at 65°C to 95°C. Such a pyrophosphatase can be readily obtained from any bacterium that is naturally able to grow and flourish at high temperatures, e.g., Thermus aquaticus. Most bacteria have naturally-occurring pyrophosphatases, and those existing in natural environments at high temperatures will therefore be suitable sources of this enzyme.

Use of a pyrophosphatase in a polymerase chain reaction as described below with Taq polymerase allows the reaction to run to completion--that is, to cause depletion of all the provided deoxynucleoside triphosphates. This allows diagnostic techniques which make use of a polymerase chain reaction to be automated. Assay for progress of the reaction can entail measurement of the generation of phosphate or the generation of DNA from the deoxynucleoside triphosphates (for example, by acid precipitation), both of which are simple and quick assays, instead of the necessity to run a gel to detect the product of the polymerase chain reaction.

- 8 -

Example 1: PCR Reaction with Pyrophosphatase

In this example DNA termed M13 Trx-F (the actual DNA used is not critical in this invention) was amplified by provision of a forward and reverse primer using a polymerase chain reaction as follows. This method is generally described in Saiki et. al., supra. Trx-F DNA at a concentration of 0.4 picomoles was mixed with 1 μ l Tris (1M, pH 8.5), 10 μ l magnesium chloride (15 mM), 6.7 μ l of four deoxynucleoside triphosphates (3 mM), 10 μ l of forward primer (10 picomole; from ALN), 20 μ l reverse primer (10 picomole, New England BioLabs), 2 μ l gelatin (0.5%), and 55 μ l distilled water. 0.5 μ l of Taq polymerase (12 units, U.S. Biochemicals, Cleveland, Ohio) was then added and the solution heated to 94°C for one minute, 50°C for one minute, and 72°C, for two minutes and this cycle of heating repeated 40 times. Identical reactions were run in the absence or presence of pyrophosphate at various concentrations (12 μ M, 37 μ M, 333 μ M, and 1 mM) and in the presence of pyrophosphatase (yeast inorganic pyrophosphatase from Sigma, Catalog No. I-4503, used without purification, or used after purification on an FPLC mono Q column). Another source of pyrophosphatase is Worthington yeast inorganic pyrophosphatase without further purification. Generally, 0.001 units of yeast inorganic pyrophosphate (4ng) are suitable in a reaction as described above. This amount may of course be considerably greater, and may be less. The range of concentrations is readily determined by routine experimentation. The concentration need only be enough to lower the level of pyrophosphate below about 5-50 μ M.

In the above reaction, pyrophosphate inhibited the polymerase chain reaction at levels of 25 μ M or

- 9 -

greater. Pyrophosphatase reversed this inhibition and stimulated production of the polymerase chain reaction products by approximately two fold.

Example 2: Preparation of Heat Resistant Pyrophosphatase

This is an example of purification of an inorganic pyrophosphatase from cells of Thermus aquaticus. Cells of T. aquaticus were obtained from the American Type Culture Collection. 10 liters of cells were grown at 70°C using the growth medium of Chien et al. 127 J. Bacteriol. 1550 (1976). The cells were harvested (~20 gm), resuspended in 40 ml of 10% sucrose, 50 mM Tris HCl, pH 7.5, 5 mM EDTA; lysed by three passages through a French press, and cell debris removed by centrifugation at 30,000 rpm, for 60 min in a Beckman 50Ti rotor. The supernatant was treated with streptomycin sulfate to remove DNA. 4 ml of a 40% streptomycin solution was added to 40 ml supernatant, mixed for 30 min., and centrifuged for 30 min at 8,000 rpm. The resulting supernatant was then treated with ammonium sulfate. No pyrophosphatase activity was precipitated at 60% ammonium sulfate, but all was precipitated by 70% ammonium sulfate: To 19 ml of supernatant 7.2 gm ammonium sulfate (60%) was added, mixed for 30 min., and spun for 30 min. at 8,000 rpm. To the supernatant 3 gm ammonium sulfate (70%) was added, mixed for 30 min., and spun for 30 min. at 8,000 rpm. The pellet was resuspended in 20 ml 20 mM Tris-HCl pH 7.5, 1 mM EDTA, 10% glycerol, 10 mM 2-mercaptoethanol (Buffer A) and then dialyzed overnight against 2 liters of Buffer A. The dialysate was passed over a DEAE DE52 column (100 ml) equilibrated in Buffer A, washed with 300 ml of Buffer A + 50 mM NaCl, and then run in a liter gradient of buffer A containing from 50 mM to 500 mM

- 10 -

NaCl. The pyrophosphatase eluted at buffer A containing 125 mM NaCl. The eluate (60 mL) was dialyzed against 2 liters of 20 mM KPO_4 pH 7.4, 1 mM EDTA, 10 mM 2-mercaptoethanol, 10% glycerol (Buffer B) and loaded onto a phosphocellulose column (100 ml) equilibrated in buffer B. All of the pyrophosphatase activity flowed through the column. This flow-through was then dialyzed against 20 mM Tris HCl pH 7.0, 1 mM EDTA, 10% glycerol (Buffer C), and applied to an FPLC monoQ column in buffer C. A gradient, in Buffer C, containing 100 mM NaCl to 250 mM NaCl was run and the pyrophosphatase activity eluted at 180 mM NaCl. Fractions with pyrophosphatase activity were dialyzed against 20 mM KPO_4 pH 7.4, 0.1 mM EDTA, 50% glycerol, and stored at -20°C .

This pyrophosphatase activity was not affected by 40 cycles of a polymerase chain reaction, with each cycle containing a 95°C , 1 min. heating step. Further, the pyrophosphatase did not hydrolyze dNTPs, nor was it inhibited by dNTPs in the reaction mixture. The pyrophosphatase activity was assayed generally as described by Chen et al. 28 Anal. Chem. 1756 (1956), and Josse, 241 J. Biol. Chem. 1938 (1966).

Other Embodiments

Other embodiments are within the following claims. For example, enzymes which use a protein primer rather than a DNA primer, e.g., $\Phi 29$ DNA polymerase which polymerizes double stranded DNA, can be used to amplify DNA without need for denaturing heating steps or reannealing steps. Blanco et al., DNA replication and mutagenesis, A.S.M. Chapter 12, 1988. Inclusion of a pyrophosphatase, or its equivalent, in such an amplification reaction will enhance the yield of DNA amplified in this system.

Claims

1 1. A solution for use in extension of an
2 oligonucleotide primer having a first single-stranded
3 region on a template molecule having a second
4 single-stranded region homologous to said first
5 single-stranded region, comprising a DNA polymerase
6 chosen from Taq polymerase, a T7-type DNA polymerase, T4
7 DNA polymerase, ϕ 29 DNA polymerase, T5 DNA polymerase,
8 and reverse transcriptase able to cause extension of
9 said first single-stranded region of said primer on said
10 second single-stranded region of said template in a
11 reaction mixture, and an agent able to reduce the amount
12 of pyrophosphate in said reaction mixture below the
13 amount produced during said extension in the absence of
15 said agent.

1 2. The solution of claim 1, wherein said
2 T7-type DNA polymerase is T7 DNA polymerase.

1 3. The solution of claim 1, wherein said
2 agent is an enzyme.

1 4. The solution of claim 3, wherein said
2 enzyme is a pyrophosphatase.

1 5. The solution of claim 4, wherein said
2 pyrophosphatase retains sufficient activity to reduce
3 the amount of pyrophosphate in said reaction mixture at
4 a temperature between 60°C and 95°C.

1 6. A kit for use in extension of an
2 oligonucleotide primer having a first single-stranded
3 region on a template molecule having a second
4 single-stranded region homologous to said first
5 single-stranded region, comprising a DNA polymerase
6 chosen from Taq polymerase, a T7-type DNA polymerase, T4
7 DNA polymerase, ϕ 29 DNA polymerase, T5 DNA polymerase,
8 and reverse transcriptase able to cause extension of
9 said second single-stranded region of said primer on
10 said second single-stranded region of said template in a
11 reaction mixture, and an agent able to reduce the amount
12 of pyrophosphate in said reaction mixture below the
13 amount produced during said extension in the absence of
14 said agent.

1 7. The kit of claim 6, wherein said T7-type
2 DNA polymerase is T7 DNA polymerase.

1 8. The kit of claim 6, wherein said agent is
2 an enzyme.

1 9. The kit of claim 8, wherein said enzyme is
2 a pyrophosphatase.

1 10. The kit of claim 9, wherein said
2 pyrophosphatase retains sufficient activity to reduce
3 the amount of pyrophosphate in said reaction mixture at
4 a temperature between 60°C and 95°C.

1 11. An improved method for extending an
2 oligonucleotide primer having a first single-stranded
3 region on a template molecule having a second
4 single-stranded region homologous to said first
5 single-stranded region, including providing a DNA

6 polymerase chosen from Taq polymerase, a T7-type DNA
7 polymerase, T4 DNA polymerase, ϕ 29 DNA polymerase, T5
8 DNA polymerase, and reverse transcriptase able to cause
9 extension of said primer on said template, the
10 improvement comprising:
11 providing an agent able to reduce the level of
12 pyrophosphate below the amount produced during said
13 extension in the absence of said agent.

1 12. The method of claim 11, wherein said
2 method further comprises the steps of providing two
3 oligonucleotide primers having first single-stranded
4 regions and two homologous template molecules having
5 second single-stranded regions homologous to said first
6 single-stranded regions, and annealing said primers to
7 said template molecules to form an annealed mixture.

1 13. The method of claim 12, wherein said
2 method further comprises the step of providing said
3 annealed mixture with said DNA polymerase and said agent
4 to cause extension of said primers.

1 14. The method of claim 12, wherein said
2 method further comprises the step of separating said
3 primers from said template molecules after said
4 extension to provide single-stranded molecules.

1 15. The method of claim 14, further comprising
2 repeating the steps of providing two primer
3 oligonucleotides, extending said primers, and separating
4 said primers.

1 16. A method for amplifying DNA comprising the
2 step of performing a polymerase chain reaction with a
3 DNA polymerase chosen from Taq polymerase, a T7-type DNA
4 polymerase, T4 DNA polymerase, ϕ 29 DNA polymerase, T5
5 DNA polymerase, and reverse transcriptase in the
6 presence of an agent able to reduce the amount of
7 pyrophosphate in said reaction below the amount produced
8 during said reaction in the absence of said agent.

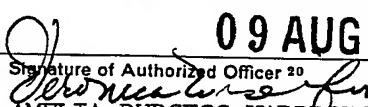
1 17. The method of claim 16, wherein said agent
2 is a pyrophosphatase.

1 18. The method of claim 16, wherein said
2 method further comprises the step of providing said
3 annealed mixture with a dideoxynucleoside triphosphate.

1 19. A method for amplifying DNA comprising
2 providing in a solution of ϕ 29 DNA polymerase and a
3 DNA to be amplified an agent able to reduce the amount
4 of pyrophosphate in said solution below the amount
5 produced in said solution in the absence of said agent.

INTERNATIONAL SEARCH REPORT

International Application No PCT/US90/01938

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all) ³ According to International Patent Classification (IPC) or to both National Classification and IPC IPC(5): C12Q 1/00; C12Q 1/68 U.S. CL.: 435/4		
II. FIELDS SEARCHED		
Minimum Documentation Searched ⁴		
Classification System	Classification Symbols	
U.S. CL.	435/4, 6, 91, 128, 183, 810, 825, 826 935/16, 78	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched ⁵		
Computer Data bases searched: U.S. Pat. and Dialog		
III. DOCUMENTS CONSIDERED TO BE RELEVANT ¹⁴		
Category *	Citation of Document, ¹⁶ with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No. ¹⁸
X	U.S., A, 4,683,202, MULLIS 28 JULY 1988 (See example 7.)	1-19
<div style="display: flex; justify-content: space-between;"> <div style="width: 45%;"> <p>* Special categories of cited documents: ¹⁵</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> </div> <div style="width: 45%;"> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"&" document member of the same patent family</p> </div> </div>		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search ² 03 JULY 1990	Date of Mailing of this International Search Report ² <div style="text-align: center; font-size: 1.2em;">09 AUG 1990</div>	
International Searching Authority ¹ ISA/US	Signature of Authorized Officer ²⁰  AMELIA BURGESS YARBROUGH	